Human neutrophil cytosolic phospholipase C: partial characterization

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Human neutrophil cytosolic phospholipase C: partial characterization

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The activity of neutrophil cytosolic phospholipase C on PIP$_2$ and PI was compared employing $[^3]$H]inositol-labeled heat-inactivated membranes of differentiated HL-60 cells, into which tracer $[^32]$P]PIP$_2$ was incorporated. Hydrolysis of PIP$_2$ did not require Ca$^{2+}$ and was stimulated when the content of PIP$_2$ in the membrane was increased by incorporation of unlabeled inositol lipid. At equal concentrations of PI and PIP$_2$ in the membrane, hydrolysis of PIP$_2$ was faster and no evidence of competition between the two substrates was obtained. Incorporation of PI into PE-$[^32]$P]PIP$_2$ vesicles, accelerated PIP$_2$ hydrolysis also at conditions that favor hydrolysis of PI. Partial purification of neutrophil cytosolic PLC on Q Sepharose, phenyl Sepharose and heparin-Agarose columns is described. From heparin-Agarose column, two PLC activity peaks exhibiting different substrate specificities were eluted. The elution profile of the main PLC species from Superose 12 gel filtration column was compatible with an approx. 150 kDa protein.

Introduction

Phosphoinositide-specific phospholipase C (PLC) participates in signal transduction initiated by hormones, neurotransmitters and growth factors [1,2]. Following cell stimulation, PLC catalyzes hydrolysis of phosphatidylinositol 4,5-bisphosphate (PIP$_2$) to generate two second messengers, sn-1,2-diacylglycerol (DAG) and inositol 1, 4, 5-trisphosphate (IP$_3$) [3]. Activation of PLC may be mediated by receptor-coupled G proteins or by the tyrosine kinase activity of receptors to growth factors [4]. Mammalian PLC in their membrane-bound or cytosolic forms consist of at least four isozymes, designated $\alpha$, $\beta$, $\tau$ and $\delta$. The isozymes differ in their structure, molecular mass and activity.

Surface-receptors for chemotactic ligands expressed by neutrophils are coupled to PLC-mediated phosphoinositide hydrolysis. Due to this, neutrophils and the related leukemia HL-60 cell line cells, have been extensively used as models for studies of signal transduction pathways involving inositol lipid turnover [5–8]. Participation of G proteins in the activation of PLC in intact and permeabilized neutrophils and in neutrophil membranes has been documented [9,10]. Only few studies, however, have been devoted to the characterization of neutrophil PLC at the molecular level [11,12]. In this present report, a partial characterization of cytosolic neutrophil PLC with respect to its activity and structure, is presented. Preliminary data of this study have been published [13].

Materials and Methods

Materials. Dowex-AG-X8 (100–200 mesh, formate form) was purchased from Bio-Rad. Culture media RPMI-1640 and M-199 were obtained from Biological Industries (Beth Haemek). All other materials were from Sigma.

Fractionation of neutrophils. Human neutrophils were isolated from fresh buffy coats by standard procedures of dextran sedimentation, Ficoll density gradient centrifugation and hemolysis. The cells were broken and fractionated as described elsewhere [14].

Synthesis of $[^32]$P]PIP$_2$ by human red blood cell PIP kinase [15]. PIP (0.6 mg/ml) was suspended by bath sonication in 50 mM Mes (pH 6.5) 2 mM EGTA/2 mM EDTA. Red blood cell membranes were suspended at 1.5 mg protein/ml in 10 mM Tris–HCl (pH
7.4) 1 mM EGTA/250 mM sucrose/0.5% CHAPS. 25 μCi of \(^{[\gamma-32P]}ATP\) (Amersham, 3000 Ci/mmol) were mixed with 100 μM ATP solution in 100 μM KP (pH 7.5) 10 mM MnCl\(_2/20\) mM MgCl\(_2\) (final volume of 10 μl). 10 μl aliquots of PIP\(_2\), ATP and red blood cell membranes were mixed and incubated at 30°C for 3.5 h. The reaction was terminated by the addition of 30 μl of 5 mM KP and 2 ml of chloroform/methanol/HCI (800:800:4, v/v). After vortexing and incubation with shaking (20 min at 37°C), 0.4 ml of 0.6 M HCl was added and phases were separated by centrifugation. The lower phase was washed twice with 1 ml chloroform/methanol/HCl (3:48:47, v/v) and its Cerenkov radiation was estimated. The radiolabeled lipid was stored at -20°C.

Preparation of PE-\(^{[32P]}\)PIP\(_2\) vesicles. Aliquots of \(^{[32P]}\)PIP\(_2\) (about 15 000 cpm) were mixed with 1 μg phosphatidylethanolamine (PE) and 2.5 μg unlabeled PIP\(_2\) dissolved in chloroform. After evaporation of the solvent under a stream of nitrogen, the lipids were suspended by bath sonication (20 min) in the reaction buffer.

Preparation of heat-inactivated, \(^{[3}H\)inositol- and \(^{[2}H\)myo-inositol-enriched membranes. HL-60 cells were labeled with \(^{[3}H\)myo-inositol (Amersham, 0.75-0.9 μCi/ml) and isolated as described [16]. The membranes were heated for 10 min at 100°C to inactivate intrinsic membrane-bound PLC. For an assay, membranes (about 15 μg of protein) were added to a test tube containing 10 000 cpm of dried \(^{[32P]}\)PIP\(_2\) and the components were sonicated in a bath sonicator (10 min) to permit incorporation of PIP\(_2\); [7]. For the preparation of PIP\(_2\)-enriched membranes, unlabeled PIP\(_2\) was pre-mixed with \(^{[32P]}\)PIP\(_2\) to give a final 1:1 molar ratio of P1:PIP\(_2\).

Enzyme activity of cytosolic PLC. Neutrophil cytosol (about 240 μg protein/ml) was incubated with one of the labeled substrates (6 min, 37°C) in 0.05 ml (for PE-PIP\(_2\) vesicles) or 0.25 ml (for labeled membranes) reaction mixtures containing 50 mM Hepes (pH 6.7) 5 mM MgCl\(_2/10\) mM LiCl/3 mM EGTA. CaCl\(_2\) was added to the desired concentration calculated according to Schatzman [18]. Activities of column fractions were determined at 2.3 μM free Ca\(^{2+}\) plus 340 μM arachidonate [16]. Hydrolysis was terminated by the addition of chloroform/methanol (2:1, v/v). Inositol phosphates were isolated on Dowex AG1-X8-formate columns eluted with 0.2 M ammonium formate/1.0 M formic acid and their radioactivities were determined in liquid scintillation counter [19].

Gel filtration of cytosolic PLC on Superose-12. Cytosol (1 mg in 0.25 ml) was fractionated on a Superose-12 column equilibrated with 10 mM KP (pH 7.0) 131 mM NaCl/0.5 mM EGTA/0.5 μM PMSF/1 μg/ml leupeptin employing an HPLC-system of Waters, Milford, MA. Fractions of 0.3 ml were collected (flow rate was 0.2 ml/min) and their enzymic activities were determined.

Chromatography of cytosolic PLC on a Q Sepharose column. Cytosol (ca. 12 mg protein) was diluted with 2 vol of 10 mM Hepes (pH 7.5) and loaded at a flow rate of 12 ml/h onto a Q Sepharose column (1.5 ml), equilibrated with 10 mM Hepes (pH 7.5) 43 mM NaCl. The column was washed and the proteins were eluted with a linear gradient of 0.043–0.6 M NaCl (20 ml). Fractions of 1 ml were collected and stored at -20°C up to 6 months without loss of activity.

Chromatography of PLC on a phenyl-Sepharose column. Active fractions from Q Sepharose were concentrated by ultrafiltration (Diaflow membrane XM-100, Amicon, MA), mixed with 0.8 M ammonium sulfate and applied (12 ml/h) to a phenyl-Sepharose column (2 ml) equilibrated with 0.8 M ammonium sulfate/10 mM KP (pH 6.7). The column was washed and proteins were eluted with 8 ml of KP-buffed 0.3 M ammonium sulfate followed by 8 ml of 40% (v/v) KP-buffed ethylene glycol. Fractions of 1 ml were collected and stored at -20°C.

Separation of cytosolic PLC forms by heparin-Agarose. Cytosol (0.4 ml, 1.65 mg protein) was diluted with an equal volume of 10 mM Hepes (pH 7.5) and applied (10 ml/h) to a column preequilibrated with 10 mM Hepes (pH 7.5) 65 mM NaCl. After washing, proteins were eluted stepwise with 0.2 M, 0.5 M and 0.8 M Hepes-buffed NaCl solutions. 1 ml fractions were collected.

Protein determination. The method of Bradford was employed, with bovine serum albumin as a standard [20].

Results

In the first part of this study, neutrophil cytosol was employed as a source of PLC to follow hydrolysis of PIP\(_2\) in PE-\(^{[32P]}\)PIP\(_2\) vesicles. Formation of the products proceeded linearly with time (up to 6 min) and with the concentration of cytosolic proteins. At concentrations exceeding about 240 μg protein/ml deviations from linearity were observed (data not shown).

PIP\(_2\) hydrolysis did not require Ca\(^{2+}\) and proceeded also in the presence of 3 mM EGTA (Table I); rates of substrate breakdown were augmented by arachidonate and Ca\(^{2+}\). Arachidonate and other cis-unsaturated fatty acids, in the presence of Ca\(^{2+}\), were previously shown to dramatically stimulate PI hydrolysis by neutrophil cytosolic PLC [16].

Incorporation of PI into the PE-\(^{[32P]}\)PIP\(_2\) vesicles at a 1:1 molar ratio to PIP\(_2\) was expected to inhibit PIP\(_2\) hydrolysis by competition between the two substrates for the available enzyme [21]. PI, however, markedly augmented rates of PIP\(_2\) breakdown, also at conditions previously shown by us to stimulate PI breakdown, i.e.,
TABLE I
The effect of PI on the hydrolysis of \[^{32}\text{P}\]PIP\(_2\) in PE-PIP\(_2\) vesicles

<table>
<thead>
<tr>
<th>Additions</th>
<th>PE-PIP(_2) vesicles</th>
<th>PE-PIP(_2)-PI vesicles</th>
</tr>
</thead>
<tbody>
<tr>
<td>None</td>
<td>1.29</td>
<td>4.16</td>
</tr>
<tr>
<td>Ca(^{2+}) (2.3 \mu M)</td>
<td>2.26</td>
<td>16.30</td>
</tr>
<tr>
<td>Arachidonate (340 \mu M)</td>
<td>1.58</td>
<td>8.56</td>
</tr>
<tr>
<td>Ca(^{2+}) plus arachidonate</td>
<td>6.41</td>
<td>22.24</td>
</tr>
</tbody>
</table>

TABLE II
PLC-catalyzed hydrolysis of \[^{31}\text{H}\]PI and \[^{32}\text{P}\]PIP\(_2\): effect of PI:PIP\(_2\) ratio

<table>
<thead>
<tr>
<th>Additions (^a)</th>
<th>Activity (pmol/min per mg protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ca(^{2+})</td>
<td>Membrane (^b)</td>
</tr>
<tr>
<td>arachid.</td>
<td>PI (^b)</td>
</tr>
<tr>
<td>-</td>
<td>(\leq 2)</td>
</tr>
<tr>
<td>-</td>
<td>(\leq 2)</td>
</tr>
<tr>
<td>+</td>
<td>(\leq 2)</td>
</tr>
<tr>
<td>+</td>
<td>165.1</td>
</tr>
</tbody>
</table>

The opposite situation, namely the effect of PIP\(_2\) on the hydrolysis of PI, was investigated in heat-inactivated membranes which may be considered closer to the natural milieu of cellular phospholipids than PE-PIP\(_2\) vesicles (see Discussion). Since labeling of PIP\(_2\) in \[^{31}\text{H}\]inositol membranes is very low \([16]\), tracer \[^{32}\text{P}\]PIP\(_2\) was incorporated into \[^{31}\text{H}\]inositol membranes to tag endogenous PIP\(_2\) \([17]\). In some of the experiments, \[^{32}\text{P}\]PIP\(_2\) was supplemented with unlabeled PIP\(_2\) to match the level of endogenous membrane PI (PIP\(_2\)-enriched membranes). Table II summarizes activities of cytosolic PLC on the two substrates in both types of membrane. In regular membranes, with PI as the predominant inositol lipid, in the absence of Ca\(^{2+}\) and arachidonate as well as in the presence of either one, rates of hydrolysis of both substrates by cytosolic PLC were low. Simultaneous presence of Ca\(^{2+}\) and arachidonate enhanced hydrolysis of both PI and PIP\(_2\), the effect on PI being much more pronounced. Enrichment of the membrane by PIP\(_2\) to a PI:PIP\(_2\) molar ratio of 1:1, substantially increased rates of PIP\(_2\) hydrolysis even in the absence of activators, indicating that in regular membranes, PIP\(_2\)-PLC operated below \(V_{max}\). Hydrolysis could be further stimulated by the addition of Ca\(^{2+}\) and arachidonate. Breakdown of PI in membranes and its stimulation by Ca\(^{2+}\) and arachi-

Fig. 1. (A) Chromatography of cytosolic PLC on a Q Sepharose column. Neutrophil cytosol (4 mg/ml, 3 ml) was applied to a Q Sepharose column (1.5 ml), equilibrated with 10 mM Hepes (pH 7.5), 43 mM NaCl and eluted with a linear gradient of NaCl. (○) \(A_{280}\), (\(\triangle\)) PI hydrolysis. In three similar experiments, the mean recovery of enzymatic activity equaled 110%. (B) Chromatography of PLC on a phenyl-Sepharose column. Active pooled fractions from Q Sepharose column were applied to a phenyl-Sepharose column (2 ml) equilibrated with 0.8 M ammonium sulfate/10 mM KP\(_4\) (pH 6.7) (a). Elution with (b) 0.3 M ammonium sulfate (c) 40% ethylene glycol. 82% of the loaded activity was recovered.
The sample loaded on the column contained 1.65 mg cytosolic proteins. The NaCl content of the eluting solutions is indicated. Hydrolysis of (a) [3H]PtdIns, (b) [32P]PtdIns, and (c) $A_{280}$. The recovery of enzymic activity was 90% with respect to both substrates.

donate were not reduced by enrichment with PtdIns in spite of simultaneous hydrolysis of the latter. Thus, no inhibition of PtdIns hydrolysis by the competing substrate, PtdIns, was observed.

GTP\gamma S (1-100 \mu M) was unable to enhance activity of cytosolic PLC when tested in either PE-PtdIns vesicles or in the double labeled membranes (Table III). This result was obtained at different concentrations of calcium ions. GDP\beta S (400 \mu M) inhibited hydrolysis of PtdIns in both forms of substrate presentation and unexpectedly NaF (10 mM) also reduced activity in both cases (Table III).

**Discussion**

Most studies on inositol lipid-specific PLC in neutrophils dealt with membrane-associated enzyme in intact, permeabilized or broken cell preparations [5–9]. The neutrophil enzyme was stimulated by binding of chemotactic ligands to surface receptors coupled to G proteins [7–10].

In a previous communication, we described a remarkable enhancement of the activity of neutrophil cytosolic PLC by unsaturated fatty acids and calcium ions [16]. Stimulating effects of anionic phospholipids and amphiphiles and their ability to relieve PtdIns hydrolysis from inhibition by long-chain phosphatidylcholine molecules, were demonstrated in other systems by Irvine et al. [22] and Hofmann and Majerus [23]. These authors attributed alterations in the activity of PLC caused by incorporation of certain lipids into the membrane, to the effects of the lipids on the physical structure of the substrate.

In the present report, we extended our earlier studies on PI-hydrolyzing activity of neutrophil cytosolic PLC [16] to hydrolysis of PtdIns in membranes and PE-PtdIns vesicles. The activity of cytosolic PLC on PtdIns in PE-PtdIns vesicles was also augmented by arachidonate and calcium; unlike PI breakdown, however, PtdIns hydrolysis proceeded also in the absence of both activating agents (Table I). The remarkable potentiation of PtdIns-hydrolyzing activity by incorporation of PI in the absence of activators, namely at conditions unfavorable for PI hydrolysis, may be attributed to alterations in the physicochemical structure of the vesicles [22–24]. Alternatively, it can be suggested that PI as well as arachidonate interact with a resting or turning-over PLC at a site distinct from the active site of the PLC.
enzyme. In a single case, binding of PLC to PI vesicles was indeed described [25].

In cellular signalling processes, PIP₂ is hydrolyzed preferentially to PI, to give two second messengers, IP₃ and diacyl glycerol [1,2]. Potentiation of PIP₂ hydrolysis by PI demonstrated in the present study (Table I), may constitute a factor that contributes to the preferential cleavage of PIP₂ in biological membranes. It should be pointed out, however, that our data were obtained employing lipid vesicles and not membranes as a substrate.

Experiments conducted with membranes (Table II) permitted direct comparison of activities of neutrophil cytosolic PLC on PI and PEP₂ in a milieu which may be considered as close as possible to the natural milieu in the cell. Although the heat pretreatment of the membranes, employed by us to inactivate intrinsic PLC, might have introduced alterations in their topography, the overall composition of the membrane was retained. Since in vivo the negatively charged inositol lipids and particularly PIP₂ are bound to membrane proteins [26], the presence of proteins in the microenvironment of the substrate may represent an advantage over pure lipid or lipid-detergent vesicles.

In HL-60 cell membranes in which the ratio of PI:PIP₂ was approx. 93.8:2.3 [27], hydrolysis of both inositol substrates by cytosolic PLC proceeded at low and similar rates (Table II). The presence of arachidonate and calcium rendered PI the preferable substrate, although cleavage of PIP₂ was also enhanced. Hydrolysis of PIP₂ was stimulated by the enrichment of the membranes with exogenous PIP₂. Assuming that the presentation of membrane-incorporated PIP₂, and intrinsic PIP₂, to the enzyme are similar [17], these data imply that prior to the enrichment, the rate of PIP₂ breakdown was below Vₘₐₓ. The experiments summarized in Table II indicate also that at equal PI and PIP₂ concentrations, neutrophil cytosolic PLC acts preferentially on PIP₂, in the presence as well as in the absence of Ca²⁺. In spite of this preferential hydrolysis of PIP₂, excess of PIP₂ incorporated into the membrane did not competitively inhibit the cleavage of PI stimulated by Ca²⁺/arachidonate (Table II). This result may be attributed to dual and opposing effects of PIP₂ acting both as a competing substrate that inhibits PI hydrolysis [21] and as a positive modulator which augments its breakdown. In membranes in which the effect of PIP₂ on PI hydrolysis was tested (Table II) these opposing effects were of similar magnitude; in the reciprocal situation represented by PE-PIP₂ vesicles enriched by PI (Table I), the positive modulation by PI was stronger resulting in a net stimulation of PIP₂ hydrolysis. An alternative explanation for the lack of mutual inhibition of PIP₂ cleavage by PI (Table I) or PI cleavage by PIP₂ (Table II) may implicate PLC isozymes that differ in their kinetic parameters (affinity, Vₘₐₓ) with respect to the two substrates. This possibility is indeed implied by chromatography on heparin-Agarose which revealed only one activity peak exhibiting substantial activity on PI (Fig. 2).

The elution profile of PLC activity from a heparin-Agarose column is consistent with the presence of at least two isozymes (Fig. 2). Most living cells contain several isozymes of PLC which differ in their structure and activity [1,2,4]. Three species of PLC were previously detected by gel filtration of neutrophil cytosol [28]. The size of one of them resembled the main PLC species of neutrophil PLC reported in this study (Fig. 4) and was compatible with the size of β or γ isozymes of PLC. PLCγ₁ which undergoes phosphorylation on tyrosyl residues by the growth factor-stimulated kinase activity of the receptor has been detected in most living cells [4]. Moreover, in the promyelocytic leukemia HL-60 cells, which upon appropriate induction undergo maturation into cells that resemble neutrophils, PLCγ and PLCβ forms have been identified [4,27].

The deviations from linearity of PLC activity observed by us at high concentrations of cytosol, cannot be attributed to exhaustion of the substrate (less than 15% was hydrolysed) nor to accumulation of products of the reaction. They may reflect the presence in the cytosol of an endogenous inhibitor loosely associated with the enzyme and its dissociation upon dilution of cytosol.

Stimulation of inositol lipid hydrolysis by guanine nucleotides compatible with involvement of G proteins in the activity of neutrophil PLC has been described by many investigators [8–10, 29–32]. In our hands, the activity of neutrophil cytosolic PLC assayed in lipid vesicles or in heat-inactivated membranes devoid of active G proteins was not affected by GTPγS (Table III). Moreover, fluoride and GDPβS exerted an unexplained inhibition of the enzymic activity (Table III). In parallel experiments carried out on membrane PLC acting on endogenous substrate, we observed stimulation of the enzyme by GTPγS (data not shown). These results may imply that either PLC isozymes present in neutrophil cytosol are not regulated by G proteins (e.g., PLCγ) or that compatible G proteins were not present in the cytosolic preparations. In the closely related permeabilized HL-60 cells, Cockcroft et al. indeed described restoration of GTPγS-elicted responses by rat brain soluble phospholipases C of β and ε types, but not by the PLCγ [26], indicating that G proteins responsible for the stimulation were present in membranes of the permeabilized cells. Contrary to this, Camps et al. [33] have recently demonstrated activation of cytosolic PLC from differentiated HL-60 cells by GTPγS, implicating soluble G protein/s in activation of the enzyme. Further characterization and resolution of neutrophil cytosolic PLC forms will be required to clarify the mode of their regulation.
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