

## Mimicking collagen *in vitro*

Collagen is the most abundant protein in many biological tissues, including the myocardial extracellular matrix (ECM), and does not only provide structural support, but also mediates many biological processes *in vivo*. For tissue engineering and disease modeling purposes human ECM is presently mimicked by animal-derived collagen. However, increasing concerns associated with animal derived materials, batch-to-batch variations, and the inability to tune the properties of collagen in a well-controlled manner result in a need for an alternative. Methacrylated Recombinant Collagen Peptide (RCP-MA, Fujifilm) is a novel collagen mimicking peptide based on human collagen I with the following characteristics:

- Based on human collagen I, containing no animal-derived components
- Highly reproducible
- Enriched with arginine-glycine-aspartic acid (RGD) sequences to control cell-adhesive properties
- Modified with methacrylic groups for well-controlled crosslinking and hydrogel fabrication in a broad range of stiffness



Figure 1: Material design for the recombinant collagen peptide based on human collagen I. Figure from CellNest data sheet (Fujifilm).

## Research Aim

In this project, we aim to explore methacrylated recombinant collagen peptide (RCP-MA, Fujifilm) as a 3D hydrogel *in vitro* platform to mimic myocardial ECM. Using this platform, we aim to assess how increasing stiffness and an increase in the number of cardiac fibroblasts, both happening upon myocardial injury, affect cardiomyocyte contractility.

## Approach

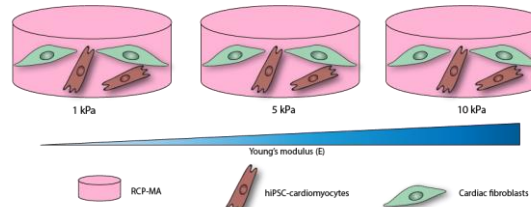


Figure 2: Graphical representation of experimental approach. Cardiomyocytes, derived from induced pluripotent stem cells (hiPSC-CMs), and human fetal cardiac fibroblasts (cFBs) are encapsulated as monoculture or co-culture inside three different types of RCP-MA hydrogels: 1) "soft" (~ 1 kPa), 2), "intermediate" (~ 5 kPa), and 3) "stiff" (~ 10 kPa). Chemical photocrosslinking is achieved using UV illumination at 5 mW/cm<sup>2</sup> and 365 nm in combination with lithium phenyl 2,4,6-trimethylbenzoylphosphinate (LAP). To assess the effect of cFB incorporation on hiPSC-CM contractility, either a monoculture of hiPSC-CMs was seeded or a co-culture representing the contractile (70% hiPSC-CM : 30% cFBs) or fibrotic (70% cFBs : 30% hiPSC-CMs) cellular environment of the myocardium.

## Results

### A: Tunability of RCP-MA stiffness

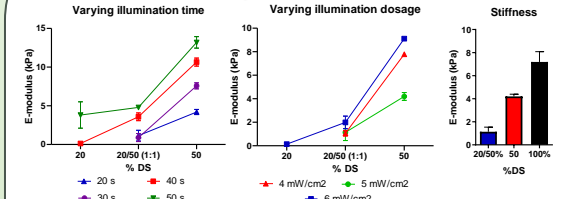


Figure 3: The effect of illumination time and dosage on the Young's modulus (E) of RCP-MA hydrogel constructs. Increasing the degree of substitution (%DS, methacrylic groups), on the RCP-MA peptides results in increasing E-modulus. Moreover, increasing the time and dosage of UV illumination results in stiffer constructs in a well-controllable manner.

### B: Cell viability and co-culture characterization

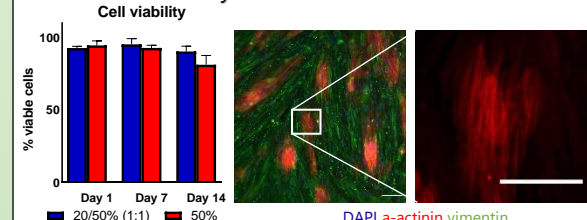


Figure 4: CFB viability and assessment of cFBs (vimentin) and hiPSC-CMs (a-actinin) phenotype encapsulated inside 20/50% RCP-MA assessed by means of immunofluorescence staining. Aligned sarcomere structures were found for hiPSC-CMs, demonstrating a maturing beating apparatus. Scale bar indicates 100 μm.

**Conclusion** Our results propose RCP-MA as an attractive alternative to animal-derived collagen for cardiac disease modeling. Taken together, these results will shed light on the importance of stiffness and cardiac fibroblast presence in achieving the physiological behavior of cardiomyocytes *in vitro*.

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